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1 **Bioavailability and toxicity of bromine and neodymium for plants grown in**  
2 **soil and water**

3

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25 **Abstract** Information about biological significance and possible phytotoxicity of many trace  
26 elements is still scarce. Bromine and neodymium are among the poorly investigated trace  
27 elements. In the research, greenhouse experiment was conducted to study effects of bromide  
28 of neodymium on wheat seedlings grown in soil and water. The wheat seedlings were capable  
29 of accumulating large amounts of both Br and Nd. Compared to the soil-grown plants, the  
30 water-grown plants accumulated higher concentrations of the trace elements. The  
31 bioaccumulation of Br and Nd resulted in statistically significant variations in the  
32 concentrations of several elements. The concentrations of P, Cl, and Ca in roots and Cl in  
33 leaves of the plants grown in the contaminated water and the concentration of I in roots of the  
34 soil-grown plants decreased. In the water-grown seedlings, the concentrations of Na and P  
35 were higher and concentrations of Mg and K were lower than those in the seedlings grown in  
36 soil. In leaves of the plants grown in water, the concentration of Cl was lower than in leaves  
37 of the soil-grown plants. In roots of the water-grown plants the concentration of Zn was  
38 higher and in leaves it was lower compare with Zn content in roots and leaves of the plants  
39 grown in soil. The K/Na ratios were 4 (leaves) and 20 (roots) times higher in the soil-grown  
40 plants, while, the Ca/Mg ratios were 8 – 19 times higher in the water-grown plants. Marked  
41 distinctions were also observed in relationships between different elements in the soil-grown  
42 and water-grown plants.

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44 **Key words** Wheat; Bromine; Neodymium; Phytoextraction from soil and water

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47 **Introduction**

48 Information about environmental chemistry of many trace elements is still scarce and often  
49 contradictory. So far, only a very short list of so-called heavy metals has attracted much

50 attention of researchers. Many other potentially toxic and/or biologically essential chemical  
51 elements often remain outside the major focus of scientists. Among these not well studied  
52 trace elements are bromine (Br) and neodymium (Nd). The role of Br and especially Nd in  
53 plant development and mineral nutrition is still poorly understood. Previously insufficient  
54 quality of analytical techniques used for determination of the elements in the environmental  
55 samples has been one of the chief causes of such a situation.

56 By 1999, the number of known natural organobromine compounds was over 1600  
57 (Gribbe 1999). Later, the number of these compounds slightly increased (Gribbe 2015). Most  
58 of the bromides have been found in marine organisms. This is due to rather high concentration  
59 of Br in the sea and because this trace element can more easily be accumulated in plants  
60 growing in water. Terrestrial plants contain much less of such compounds.

61 The reported range of Br concentrations in plants growing in non-contaminated by  
62 bromides soils varies from  $<1 \text{ mg kg}^{-1}$  to  $30 \text{ mg kg}^{-1}$  (Tensho 1970; Låg and Steinnes 1977;  
63 Gan et al. 1998; Wishkerman 2006; Sahin et al. 2012; Pourimani et al. 2013; Kabata-Pendias  
64 and Szteke 2015; Shtangeeva et al. 2017). Previously methyl bromide was the most well-  
65 known Br compound. It was used as a fumigant to regulate the soil-borne pesticides  
66 (Schneider et al. 2003). Although methyl bromide demonstrated a good effectiveness, it was  
67 found that the active use of the fumigant leads to depletion of the ozone layer (Shorter et al.  
68 1995). Owing to this, the use of methyl bromide was limited in many countries. However, at  
69 present, numerous man-made Br compounds are widely used and released into the  
70 environment. Since these compounds may be toxic, it is important to study distribution of  
71 different bromides in soils and plants and estimate the fate of the Br bioaccumulation.  
72 Unfortunately, up to now, there is not enough available information on concentrations of Br in  
73 different plant species.

74 Among other lanthanides, there is much less information on Nd in the environment. This  
75 trace element is one of the most critical rare earth elements until 2025 (Freitas et al. 2020).  
76 One of the main applications of Nd is Nd-Fe-B magnets that are widely used in computers,  
77 cell phones and many other components (München and Veit 2017). The biological role of Nd  
78 is currently unknown. It is assumed that Nd is moderately toxic trace element (Zhao et al.  
79 2019). However, it is possible that in the long run the accumulation of Nd in the environment  
80 may have severe effects.

81 According to available reports, the range of the Nd concentration in non-contaminated  
82 soils is 5.8-53 mg kg<sup>-1</sup> (Tyler and Olsson 2002; Masto et al. 2011; Carpenter et al. 2015; Patra  
83 et al. 2020; Turra et al. 2020; Wang et al. 2020). In terrestrial plants growing in non-  
84 contaminated soils, the concentration of Nd can vary from 0.10 mg kg<sup>-1</sup> to 2.0 mg kg<sup>-1</sup> (Tyler  
85 2004; Carpenter et al. 2015; Romero-Freire et al. 2019; Gorena et al. 2020). Although  
86 possible phytotoxicity of Nd is still poorly understood, it may be assumed that the toxic  
87 effects will depend on the concentration of the trace element in growth medium. It has been  
88 reported that at low doses Nd can have a stimulatory effect on the plant development (Luo et  
89 al. 2008; Rezaee et al. 2018) and reduce toxicity of well-known toxicants such as Cr (Lu et al.  
90 2020).

91 The accumulation of elements in soil-grown plants is influenced by many different  
92 factors. This can make further interpretation of experimental results difficult. The process of  
93 uptake of elements by plants growing in water media is simpler. Due to this, until now, most  
94 of the experiments on phytoextraction of macro- and trace elements are usually carried out  
95 hydroponically. Meanwhile, it can be expected that the mechanisms controlling transfer of  
96 elements from growth medium to plants can differ when the plants grow in different media. In  
97 water, mineral elements are present in more available to a plant form and can be easily taken  
98 by roots. In soil, a certain part of an element is often adsorbed onto surface of the soil

99 particles. Therefore, even though concentration of the element in the soil and water is the  
100 same, the roots will take from the growth media different amounts of the element. It may be  
101 assumed that translocation of elements from roots to upper plant parts will also differ in the  
102 water-grown and soil-grown plants.

103 Based on these assumptions, we studied effects of bromide of Nd on the plants grown  
104 under different conditions – in soil and in water. We performed the experiment when plants  
105 grow in non-contaminated soil and water and in the media contaminated by bromide of Nd.  
106 The research was addressed to a deeper understanding the Br and Nd pathways in plants and  
107 evaluation of the factors important for bioaccumulation of the trace elements. The main aims  
108 of the experiment were the following:

- 109 (1) to study the potential of wheat to phytoextract Br and Nd from different growth media  
110 (soil and water);
- 111 (2) to assess the effects of the Br and Nd bioaccumulation on the concentrations of other  
112 elements in the plants;
- 113 (3) to examine the uptake of macro- and trace elements and to compare the relationships  
114 between the elements in the plants grown in soil and in water.

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## 117 **Materials and Methods**

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119 Experimental design

120

121 Eighty seeds of wheat *Triticum aestivum* L. were rinsed several times by ultra-pure water and  
122 germinated on a moist filter paper during five days. The five-day-old germinated seedlings  
123 were divided into two parts. One part of the seedlings was transferred to pots volume of 5 kg

124 filled with soil. The soil had a loamy sand texture (74% sand, 24% silt, 2% clay). Second part  
125 of the seedlings was placed into vessels volume of 600 mL filled with tap water. Each series  
126 of the experiment consisted of two parts: one half of the seedlings were grown in non-  
127 contaminated media and second part was grown in the water or in the soil spiked with  
128 bromide of Nd ( $\text{NdBr}_3 \times 6\text{H}_2\text{O}$ ). The concentration of Br in the solution was  $50 \text{ mg L}^{-1}$ . The  
129 wheat seedlings were grown in a naturally illuminated greenhouse during 10 days. A  
130 completely randomized design was used. The pots were watered daily. The air temperature  
131 was usually  $25^\circ\text{C}$  during the day and  $22^\circ\text{C}$  at night. The experiment was performed in  
132 triplicate. After sampling, the plants were carefully washed by deionized water and air-dried  
133 up to constant weight.

134

135 Elemental analysis

136

137 The extraction procedure used in the research was optimized in our earlier studies  
138 (Shtangeeva et al. 2015; Shtangeeva et al. 2017). In this procedure, the dried roots and leaves  
139 of wheat seedlings and reference material were weighed into the 50 ml conical centrifuge  
140 tubes (Falcon). Two ml of tetramethyl ammonium hydroxide (TMAH) solution were added to  
141 the vessels. Then the vessels were closed, and samples were heated during 16 hours in a sand  
142 bath at  $60^\circ\text{C}$ . After extraction, the samples were diluted to 25 ml with ultrapure water. Before  
143 elemental analysis, the samples were stored in a refrigerator at a temperature of  $+5^\circ\text{C}$ . The  
144 ICP-OES (Agilent 5110 VDV) was used for determination of Na, Mg, P, K, Zn, and Mn. The  
145 concentrations of Cl, Br, I, and Nd were determined by ICP-MS. For the ICP-OES  
146 determination, all the samples were diluted to 1:2 with 5% (v/v)  $\text{HNO}_3$ ; for the ICP-MS  
147 analysis, the samples were diluted 1:2 with ultrapure water. Neodymium ( $^{146}\text{Nd}$ ) was  
148 measured by Agilent 8900 ICP-MS/MS. Furthermore, a Thermo Elemental X7 quadrupole

149 ICP-MS was used in the determination of  $^{37}\text{Cl}$ ,  $^{81}\text{Br}$  and  $^{127}\text{I}$  (Shtangeeva et al. 2015). The ion  
150 lens settings, nebulizer gas flow rate and torch position of the instrument were optimized to  
151 obtain the maximum  $^{115}\text{In}$  count rate. To avoid matrix effects in sample introduction, halogens  
152 were measured from alkaline solutions (4% (v/v) TMAH). In addition, 4% TMAH (v/v)  
153 solution was used as a washing solution between the samples when halogens were measured.

154

#### 155 Quality Control

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157 For control of the quality of the analytical measurements was used standard reference material  
158 (SRM) Tomato leaves 1573a (National Institute of Standards and Technology, USA). The  
159 differences between the certified and informative values and concentrations of elements  
160 determined in the SRM in our work did not exceed 5-10%.

161

#### 162 Data analysis

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164 The multivariate statistical analysis of experimental results was carried out using Statistica for  
165 Windows 6.0 Software packages (StatSoft, Tulsa, OK, USA). Before analysis, the normality  
166 of the distribution of the analytical data was checked by the Shapiro-Wilk test. We calculated  
167 mean concentrations of elements and performed analysis of variances in order to assess  
168 statistically significant ( $P < 0.05$ ) differences between samples. Pearson correlation analysis  
169 and cluster analysis were used to assist in the identifying the factors that could influence on  
170 the bioaccumulation of elements and on the interactions between the elements in plants. This  
171 information can help in a better understanding the mechanisms that affect the uptake of  
172 elements and assess the relationships between elements that are typical for the plants growing

173 in different media, including when the growth media are non-contaminated or contaminated  
174 by bromide of Nd.

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## 177 **Results and discussion**

178

179 Bromine and Nd accumulation in wheat seedlings grown in soil and water spiked with  
180 bromide of Nd

181

182 As a result of growth of wheat seedlings in the soil and water contaminated by  $\text{NdBr}_3 \times 6\text{H}_2\text{O}$   
183 the concentration of Br in the plants increased (Table 1). The changes in the Br concentrations  
184 were different for the plants grown in water and soil. The concentrations of Br in roots and  
185 leaves of the water-grown plants increased much more than in roots and leaves of the plants  
186 grown in soil. The transfer of Br from roots to leaves also differed. In leaves of the water-  
187 grown wheat seedlings, the Br concentration was almost 2 times lower than in roots. In the  
188 soil-grown plants, the Br concentration in leaves was 1.5 times higher as compared to the Br  
189 content in roots.

190 Compared to control, the concentration of Nd in the wheat seedlings grown in the  
191 contaminated media increased significantly. A greater increase was observed in the plants  
192 grown in water medium. Roots accumulated more Nd as compared to leaves. The higher  
193 accumulation of Br and Nd in roots and leaves of the water-grown plants can be due to a  
194 better availability of nutrients and also non-essential trace elements to the plants grown in  
195 water media. It was reported that usually a large part of trace elements can be adsorbed onto  
196 roots, and only a small part of the elements will be transferred to upper plant parts (Chandra et  
197 al. 2009; Klink et al. 2013). Based on this assumption, we could expect lower Br and Nd



198 concentrations in leaves as compared to concentrations of the elements in roots (when the  
199 plants grow in the medium enriched with bromide of Nd). In fact, this was observed for Nd,  
200 but in the case of Br, it was only found in the water-grown plants. When the wheat seedlings  
201 were grown in the Br-contaminated soil, the translocation of Br from roots to leaves was  
202 enhanced. In our previous experiment (Shtangeeva et al. 2017) we studied accumulation of Br  
203 in pea and wheat seedlings grown in the soils spiked with KBr and NaBr and also found that  
204 concentration of Br in leaves of wheat was always higher than in roots. On the other hand,  
205 transfer of this trace element from roots to leaves of pea was suppressed. Similar distribution  
206 of Br between roots and leaves of wheat seedlings was also observed by other researchers  
207 (Zhang et al. 2013; Zhu et al. 2020). One of the possible explanations for this phenomenon  
208 could be the following. Although the uptake of nutrients and trace elements by plants is  
209 affected by physical, chemical and biological soil characteristics (Chapin et al. 1987), the  
210 concentrations of elements in one or another plant species are determined by phylogeny of the  
211 plants (Watabene et al. 2007). That is why we often can observe quite different processes of  
212 element uptake and translocation from roots to upper plant parts in different plant species  
213 grown simultaneously and under the same conditions.

214

215 Variations in the concentrations of other elements in wheat seedlings resulting from  
216 bioaccumulation of Br and Nd

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218 The growth of wheat seedlings in soil and water spiked with bromide of Nd also affected  
219 concentrations of some other elements in the plants. As expected, more serious effects were  
220 found in roots, especially in roots of the water-grown plants. Compared to control, the  
221 concentrations of P, Cl, and Ca in roots of the plants grown in the contaminated water  
222 decreased statistically significantly ( $P < 0.05$ ). In roots of the soil-grown plants such a decrease

223 was found only for I. There were no statistically significant changes in the concentrations of  
224 elements in leaves of the soil-grown plants. In leaves of the plants grown in the water  
225 contaminated by  $\text{NdBr}_3 \times 6\text{H}_2\text{O}$  we observed a statistically significant ( $P < 0.05$ ) variation only  
226 for Cl.

227 Thus, plants grown in water were more affected by  $\text{NdBr}_3 \times 6\text{H}_2\text{O}$  contamination than  
228 plants grown in soil. It may be assumed that main effect on the decrease of halogens might be  
229 caused by bioaccumulation of Br (result of competition between chemically similar elements  
230 during their uptake by plants). An increase of Nd concentration in the seedlings could  
231 additionally lead to the decrease of P and Ca contents in the plants grown in the bromide  
232 contaminated water. The similar ionic radii of Nd and Ca suggest that Nd can compete with  
233 this essential plant nutrient. It was shown that Nd(III) can replace Ca(II) in plants (Wei et al.  
234 2001; Zhang and Shan 2001). Rare earth phosphate is main component of rare earth ore  
235 (Onoda et al. 2002). Therefore, the relationship between Nd (rare earth element) and P is  
236 expectable.

237

238 Differences in the elemental composition of water-grown and soil-grown plants

239

240 Concentrations of almost all macro- and trace elements in the plants grown in soil and water  
241 differed significantly ( $P < 0.05$ ). In roots and leaves of the wheat seedlings grown in non-  
242 contaminated and contaminated water the concentrations of Na and P were higher and  
243 concentrations of Mg and K were lower than those in the seedlings grown in soil. Besides, in  
244 roots of the water-grown plants, the concentration of Zn was higher and in leaves it was lower  
245 in comparison with Zn content in roots and leaves of the plants grown in soil. In leaves of the  
246 plants grown in non-contaminated water, the concentration of Cl was lower and concentration  
247 of Br was higher as compared to concentrations of the elements in leaves of the soil-grown

248 plants. In roots, the concentrations of both these elements were almost the same. In previous  
249 experiment, a higher accumulation of Br was also observed in leaves of the wheat grown in  
250 non-contaminated with Br liquid media (distilled water, spring water, nutrient solution of  
251 Hoagland) as compared to accumulation of Br in leaves of the wheat grown in soil  
252 (Shtangeeva 2017). One may speculate that the higher transfer of Br from roots to leaves of  
253 wheat is due to distinctive physiological characteristics of the plant species.

254 It is well-known that K and Mg play a vital part in plant development. Potassium is main  
255 inorganic cation in the cytoplasm of plants. This nutrient is essential for proper function of  
256 many enzymes (Marschner 1995). Magnesium is crucial in biosynthesis of chlorophyll and  
257 carbon fixation (Guo et al. 2015). The decrease in uptake of K and Mg implies that growth of  
258 plants in water may directly interfere with plant growth, development, and photosynthesis.

259

260 Ratios of essential nutrients in the plants grown in soil and water

261

262 Table 2 shows the ratios of essential nutrients in the plants grown in control and contaminated  
263 by bromide of Nd water and soil. The K/Na ratios were much higher in leaves and especially  
264 in roots of the soil-grown plants as compared with the K/Na ratios in the plants grown in  
265 water. On the other hand, the Ca/Mg ratios were higher in the water-grown plants than in the  
266 plants grown in soil. There was no effect of contamination of the growth media by bromide of  
267 Nd on the K/Na ratios in the plants grown both in soil and in water. In the soil-grown plants,  
268 the Ca/Mg ratios also did not change as a result of contamination of the growth medium.  
269 However, as compared to the wheat seedlings grown in clean water, the Ca/Mg ratios in the  
270 seedlings grown in the contaminated water were lower in roots and higher in leaves. As it was  
271 suggested, the toxic action of Nd on plants may include replacement of Ca/Mg (Sneller et al.  
272 2000).

273

274 Statistical analysis of experimental data

275

276 Figure 1 shows results of cluster analysis of the wheat seedlings grown in non-contaminated  
277 and contaminated by  $\text{NdBr}_3 \times 6\text{H}_2\text{O}$  media. Roots and leaves of the plants grown in soil were  
278 well separated into two groups (Fig. 1a). Moreover, in each group there was a good separation  
279 of the plants grown in clean soil and in the soil spiked with bromide. In the water-grown  
280 seedlings, the separation between roots and leaves, as well as between plants grown in non-  
281 contaminated and contaminated media, was less noticeable. (Fig. 1b).

282 The results of the correlation analysis clearly indicate the differences in the relationships  
283 between elements in the plants grown in the different media. In leaves of the soil-grown  
284 plants, the correlation between Mg and Ca – chemically similar elements – was statistically  
285 significant and positive ( $r = 0.95$ ). In leaves of the plants grown in water, the correlation  
286 between these elements was also statistically significant but negative ( $r = -0.82$ ). The  
287 correlation between concentrations of Ca in roots and leaves of the soil-grown plants was  
288 negative ( $r = -0.83$ ,  $P < 0.05$ ). The statistically significant negative correlation was also found  
289 between concentrations of K in roots and leaves of the wheat seedlings grown in soil ( $r = -$   
290  $0.71$ ). However, there was no correlation between concentrations of Ca (and also between  
291 concentrations of K) in roots and leaves of the water-grown plants. This fact can serve as  
292 additional evidence that mechanisms of accumulation of elements in the plants grown in soil  
293 and in water media are different. Therefore, there is reasonable to think that it is hardly  
294 possible to use experimental data on accumulation of different elements in the plants grown  
295 hydroponically in order to appreciate the mechanisms of the element uptake by the plants  
296 growing in natural conditions (in soil).

297

298

299 **Conclusions**

300 Our experimental results showed that growth of wheat seedlings in the media contaminated  
301 by bromide of Nd resulted in an increase of Br and Nd concentrations in the plants. The  
302 greatest accumulation was found in the water-grown plants. Compared to control, the  
303 concentrations of Br and Nd in the plant roots increased 215 and 340 times, respectively,  
304 while in the soil-grown wheat seedlings, the increase of Br and Nd plant concentrations was  
305 10 times less. The bioaccumulation of these potentially toxic trace elements led to variations  
306 in the concentrations of several other elements. Plants grown in contaminated water medium  
307 showed statistically significant changes in the concentrations of P, Cl, and Ca (roots) and I  
308 (leaves). In the soil-grown plants, such a variation was observed only for I in roots. The  
309 concentrations of many elements (Na, Mg, P, K, Ca, and Zn) in roots and leaves of the water-  
310 grown plants differed significantly as compared to concentrations of the elements in the plants  
311 grown in soil.

312

313

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318

319

320 **Compliance with Ethical Standards**

321 The authors have no conflicts of interest to declare that are relevant to the content of this  
322 article. This article does not contain any studies involving animals or human participants  
323 performed by any of the authors.

324

325

326 **Data Availability**

327 The authors confirm that the data supporting the findings of this study are available within the  
328 article.

329

330

331 **Contribution of authors to the manuscript**

332 Study conception and design: Irina Shtangeeva; analysis of experimental samples: Matti  
333 Niemelä, Paavo Perämäki; draft manuscript preparation: Irina Shtangeeva. All authors  
334 reviewed the results and approved the final version of the manuscript.

335

336

337 **Consent to Participate and Consent to Publish**

338 Not applicable to the manuscript.

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340

341 **References**

342 Carpenter, D., Boutin, C., Allison, J. E., Parsons, J. L., & Ellis, D. M. (2015). Uptake and  
343 effects of six rare earth elements (REEs) on selected native and crop species growing in  
344 contaminated soils. *Plos one*, <https://doi.org/10.1371/journal.pone.0129936>.

345 Chandraa, R., Bharagavaa, R. N., Yadava, S., & Mohan, D. (2009). Accumulation and  
346 distribution of toxic metals in wheat (*Triticum aestivum* L.) and Indian mustard (*Brassica*  
347 *campestris* L.) irrigated with distillery and tannery effluents. *Journal of Hazardous*  
348 *Materials*, 162, 1514–1521.

349 Chapin, F. S., Bloom, A. J., Field, C. B., & Waring, R. H. (1987). Plant responses to multiple  
350 environmental factors. *BioScience*, 37(1), 49-57.

351 Freitas, R., Costa, S., Cardoso, C. E. D., Morais, T., Moleiro, P., Matias, A. C., et al. (2020).  
352 Toxicological effects of the rare earth element neodymium in *Mytilus galloprovincialis*.  
353 *Chemosphere*, <https://doi.org/10.1016/j.chemosphere.2019.125457>.

354 Gan, J., Yates, S. R., Ohr, H. D., & Sims, J. J. (1998). Production of methyl bromide by  
355 terrestrial higher plants. *Geophysical Research Letters*, 25(19), 3595-3598.

356 *macrocarpa* leaves for biomonitoring the environmental impact of an industrial complex: The  
357 case of Puchuncaví-Ventanas in Chile. *Chemosphere*,  
358 <https://doi.org/10.1016/j.chemosphere.2020.127521>.

359 Guo, W., Chen, S., Hussain, N., Cong, Y., Liang, Z., & Chen K. (2015). Magnesium stress  
360 signaling in plant: Just a beginning, *Plant Signaling & Behavior*, DOI:  
361 10.4161/15592324.2014.992287.

362 Gribble, G. W. (1999). The diversity of naturally occurring organobromine compounds.  
363 *Chemical Society Reviews*, 28, 335–346.

364 Gribble, G. W. (2015). A recent survey of naturally occurring organohalogen compounds.  
365 *Environmental Chemistry*, 12(4), 396-405.

366 Kabata-Pendias, A., & Szteke, B. (2015). *Trace elements in abiotic and biotic environments*.  
367 London-New York: CRC Press Taylor & Francis Group.

368 Klink, A., Macioł, A., Wisłocka, M., & Krawczyk, J. (2013). Metal accumulation and  
369 distribution in the organs of *Typha latifolia* L. (cattail) and their potential use in  
370 bioindication. *Limnologica*, 43(3), 164-168.

371 Lu, N. H., Wu, L. M., Yang, R., Li, H., & Shan, C. J. (2020). Neodymium improves the  
372 activity of ascorbate-glutathione cycle and chloroplast function of wheat seedlings under  
373 chromium stress. *Photosynthetica*, 58(3), 748-754.

374 Luo, J., Zhang, J., & Wang, Y. (2008). Changes in endogenous hormone levels and redox  
375 status during enhanced adventitious rooting by rare earth element neodymium of  
376 *Dendrobium densiflorum* shoot cuttings. *Journal of Rare Earths*, 26(6), 869-874.

377 Låg, J., & Steinnes, E. (1977). Halogens in barley and wheat grown at different locations in  
378 Norway. *Acta Agriculturae Scandinavica*, 27(4), 265-268.

379 Marschner, H. (1995). *Mineral nutrition of higher plants*. London, UK: Academic Press Ltd.

380 Mastro, R. E., Ram, L. C., Verma, S. K., Selvi, V. A., George, J., Tripathi, R. C., et al. (2011).  
381 Rare earth elements in soils of Jharia coal field. *International Journal of Geological and*  
382 *Environmental Engineering*, 5, 653-658.

383 München, D. D., & Veit, H. M. (2017). Neodymium as the main feature of permanent  
384 magnets from hard disk drives (HDDs). *Waste Management*, 61, 372-376.

385 Onoda, H., Nariai, H., Moriwaki, A., Maki, H., & Motooka, I. (2002). Formation and catalytic  
386 characterization of various rare earth phosphates. *Journal of Materials Chemistry*,  
387 12(6), 1754–1760.

388 Patra, A. C., Lenka, L., Sahoo, S. K., Jha, S. K., & Kulkarni, M. S. (2020). Probing rare earth  
389 element distributions in soils of the mineralized Singhbhum region in India using INAA.  
390 *Applied Radiation and Isotopes*, <https://doi.org/10.1016/j.apradiso.2020.109360>.



391 Pourimani, R., Abasnejad, K., Ghanbarzadeh, K., Reza Zare, M., & Kamali, M. (2013).  
392 Determining the amount of Br, Na and K in six wheat samples with neutron activation  
393 analysis (NAA) method in Arak, I.R. Iran. *Journal of Radioanalytical Nuclear*  
394 *Chemistry*, 295, 163–166.

395 Rezaee, A., Hale, B., Santos, R. M., & Chiang, Y. W. (2018). Accumulation and toxicity of  
396 lanthanum and neodymium in horticultural plants (*Brassica chinensis* L. and *Helianthus*  
397 *annuus* L.). *Canadian Journal of Chemical Engineering*, 96(10), 2263-2272.

398 Romero-Freire, A., Turlin, F., André-Mayer, A-S., Pelletier, M., Cayer, A., & Giamberini, L.  
399 (2019). Biogeochemical cycle of lanthanides in a light rare earth element-enriched  
400 geological area (Quebec, Canada). *Minerals*, <https://doi.org/10.3390/min9100573>.

401 Sahin, O., Taskin, M. B., Kadioglu, Y. K., Inal, A., Gunes, A., & Pilbeam, D. J. (2012).  
402 Influence of chloride and bromate interaction on oxidative stress in carrot plants. *Scientia*  
403 *Horticulturae*, 137, 81–86.

404 Schneider, S. M., Roskopf, E. N., Leesch, J. G., Chellemi, D. O., Bull, C. T., & Mazzola, M.  
405 (2003). United States Department of Agriculture-Agricultural Research Service research  
406 on alternatives to methyl bromide: pre-plant and post-harvest. *Pest Management Science*,  
407 59(6-7), 814-26.

408 Shorter, J. H., Kolb, C. E., Crill, P. M., Kerwin, R. A., Talbot, R. W., Hines, M. E., et al.  
409 (1995). Rapid degradation of atmospheric methyl bromide in soils. *Nature*, 377(6551),  
410 717-719.

411 Shtangeeva, I., Niemelä, M., Perämäki, P., & Timofeev, S. (2015). Response of wheat and  
412 pea seedlings on increase of bromine concentration in the growth medium.  
413 *Environmental Science and Pollution Research*, 22, 19060–19068.

414 Shtangeeva, I., Niemelä, M., Perämäki, P., Ryumin, A., Timofeev, S., Chukov, S., et al.  
415 (2017). Phytoextraction of bromine from contaminated soil. *Journal of Geochemical*  
416 *Exploration*, 174, 21–28.

417 Shtangeeva, I. (2017). Bromine accumulation in some crops and grasses as determined by  
418 neutron activation analysis. *Communications in Soil Science and Plant Analysis*, 48(19),  
419 2338-2346.

420 Sneller, F. E. C., Kalf, D. F., Weltje, L., & Van Wezel, A. P. (2000). Maximum permissible  
421 concentrations and negligible concentrations for rare earth elements (REEs). National  
422 Institute of Public Health and Environmental Protection RIVM, Bilthoven (Netherlands).  
423 [https://inis.iaea.org/search/search.aspx?orig\\_q=RN:31053152](https://inis.iaea.org/search/search.aspx?orig_q=RN:31053152).

424 Tensho, K. (1970). Iodine and bromine in soil-plant system with special reference to  
425 "reclamation-akagare disease" of lowland rice. *Japan Agricultural Research Quarterly*,  
426 5(3), 26-32.

427 Turra, C., De Nadai Fernandes, E. A., Bacchi, M. A., Sarriés, G. A., Reyes, A. E. L. (2020).  
428 Temporal variability of rare earth elements in Ultisol soil under citrus plants. *Journal of*  
429 *Radioanalytical and Nuclear Chemistry*, 324, 219–224.

430 Tyler, G., & Olsson, T. (2002). Conditions related to solubility of rare and minor elements in  
431 forest soils. *Journal of Plant Nutrition and Soil Science*, 165, 594–601.

432 Tyler, G. (2004). Rare earth elements in soil and plant systems – A review. *Plant and Soil*,  
433 267, 191–206.

434 Wang, L., Christakos, G., Wu, C., & Wu, J. (2020). Spatial variability assessment of La and  
435 Nd concentrations in coastal China soils following 1000 years of land reclamation.  
436 *Journal of Soils and Sediments*, 20, 1651–1661.

437 Watanabe, T., Broadley, M.R., Jansen, S., White, P. J., Takada, J., Satake, K., et al.  
438 (2007). Evolutionary control of leaf element composition in plants. *New Phytologist*,  
439 174(3), 516-523.

440 Wei, Z. G., Yin, M., Zhang, X., Hong, F. S., Li, B., Tao, Y., Zhao, G. W., Yan, C. H. (2001).  
441 Rare earth elements in naturally grown fern *Dicranopteris linearis* in relation to their  
442 variation in soils in South-Jiangxi region (Southern China), *Env. Pol.* 114, 345–355.

443 Wishkerman, A. (2006). Bromine and iodine in plant-soil systems. Dissertation, Universität  
444 Heidelberg.

445 Zhang, S., & Shan, X. (2001). Speciation of rare earth elements in soil and accumulation by wheat  
446 with rare earth fertilizer application. *Environmental Pollution*, 112(3), 395-405.

447 Zhang, Y., Sun, H., Liu, F., Dai, Y., Qin, X., Ruan, Y., et al. (2013).  
448 Hexabromocyclododecanes in limnic and marine organisms and terrestrial plants from  
449 Tianjin, China: Diastereomer- and enantiomer-specific profiles, biomagnification, and  
450 human exposure. *Chemosphere*, 93(8), 1561-1568.

451 Zhao, C.-M., Shi, X., Xie, S.-Q., Liu, W.-S., He, E.-K., Tang, Y.T., & Qiu, R.-L. (2019).  
452 Ecological risk assessment of neodymium and yttrium on rare earth element mine sites in  
453 Ganzhou, China. *Bulletin of Environmental Contamination and Toxicology*, 103, 565–  
454 570.

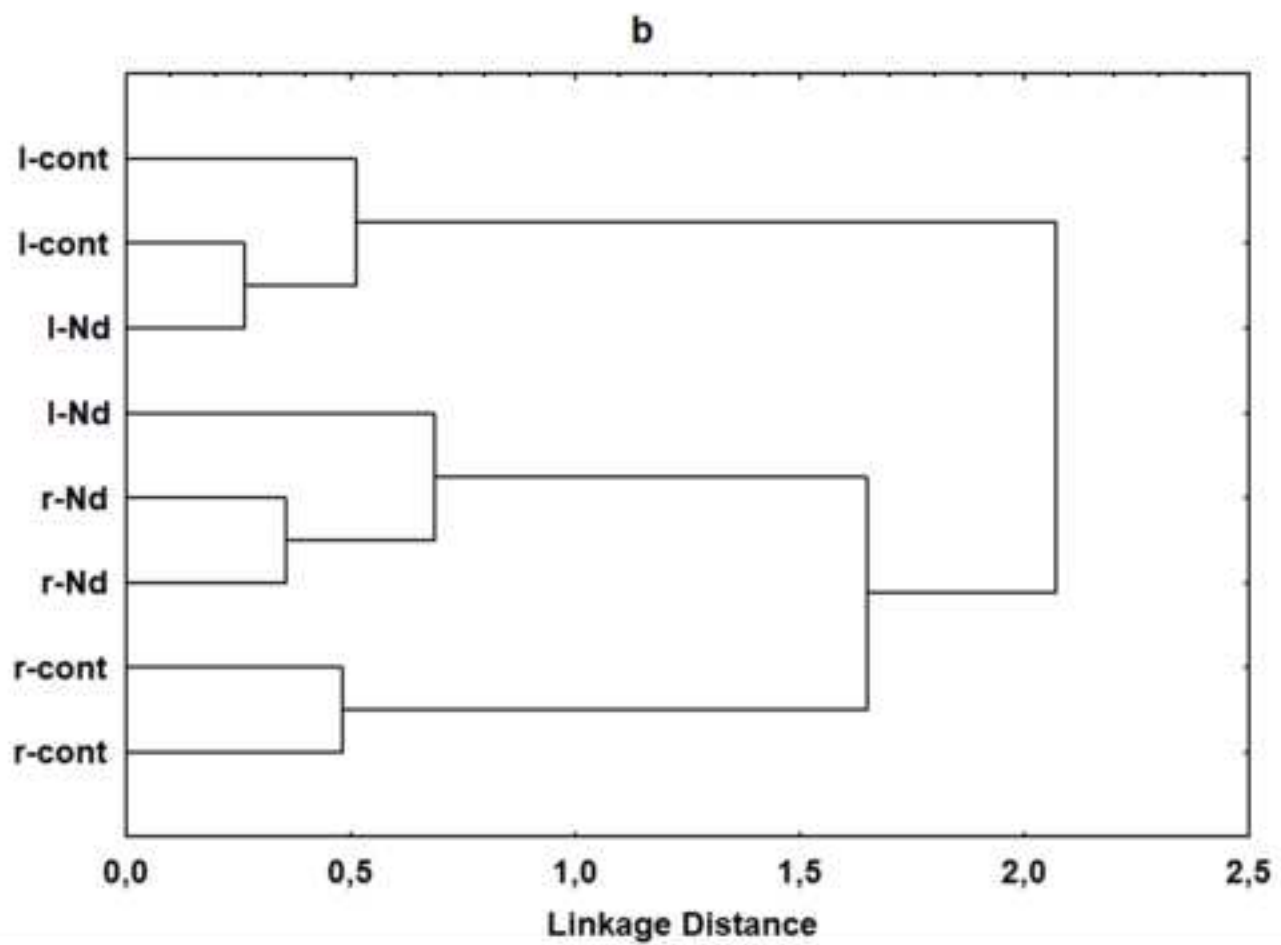
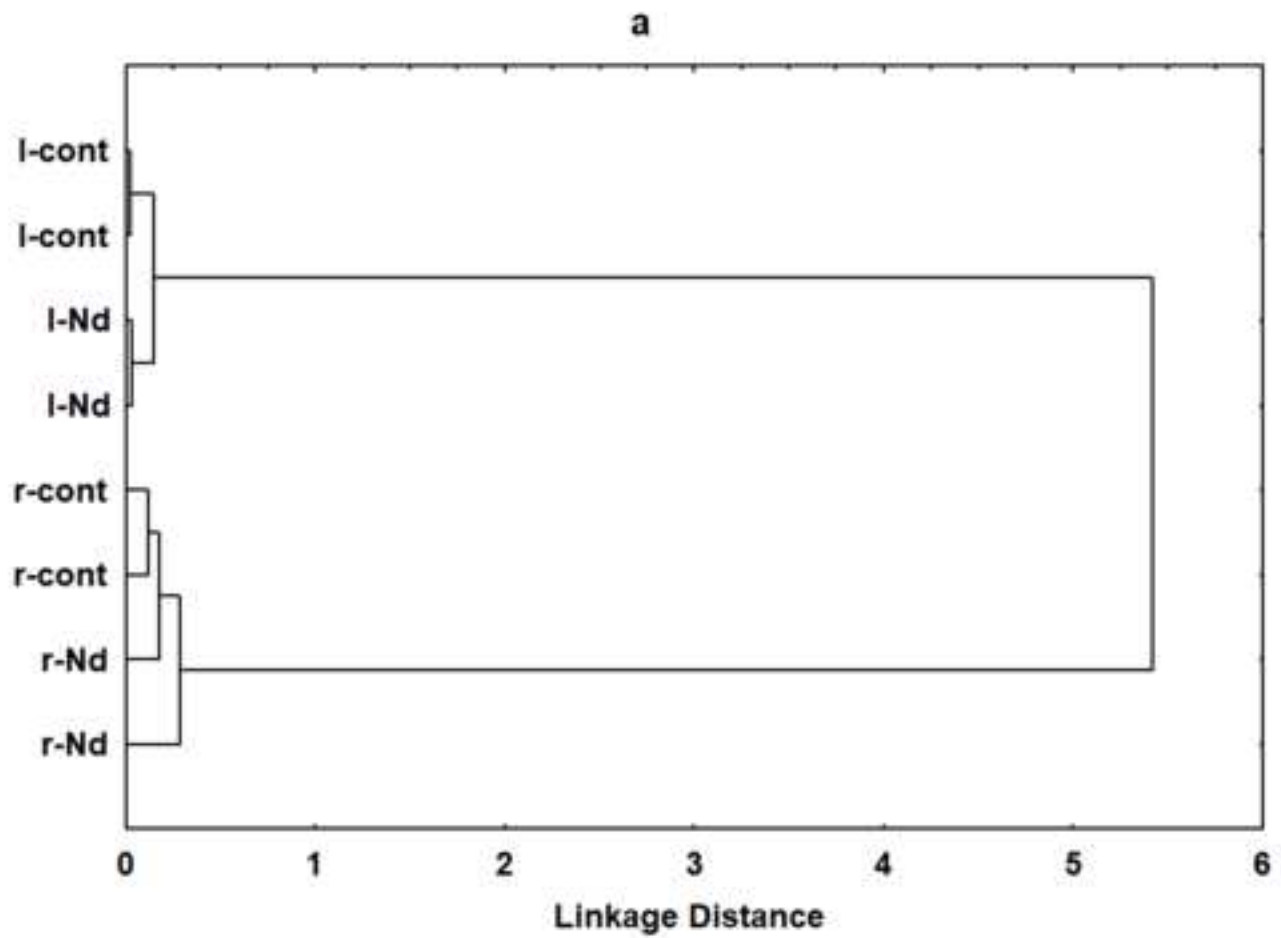
455 Zhu, H., Wang, F., Li, B., Yao, Y., Wang, L., & Sun, H. (2020). Accumulation and  
456 translocation of polybrominated diphenyl ethers into plant under multiple exposure  
457 scenarios. *Environment International*, <https://doi.org/10.1016/j.envint.2020.105947>.

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462 **Figure caption**

463 **Fig. 1** Cluster analysis (Ward's method) of soil-grown (a) and water-grown (b) wheat  
464 seedlings.

465 l - leaves, r – roots of the seedlings grown in clean media (cont) and in the media spiked with  
466 bromide of Nd (Nd)



**Table 1** Mean concentrations (n=3)  $\pm$  SD of elements in wheat seedlings.1 – control, 2 - the growth medium was spiked with  $\text{NdBr}_3 \times 6\text{H}_2\text{O}$ 

Element	Roots (water-grown plants)		Roots (soil-grown plants)	
	1	2	1	2
Na, %	3.6 $\pm$ 0.4 <sup>a</sup>	3.2 $\pm$ 0.3 <sup>a</sup>	0.27 $\pm$ 0.04	0.27 $\pm$ 0.03
Mg, %	0.41 $\pm$ 0.04 <sup>a</sup>	0.53 $\pm$ 0.04 <sup>a</sup>	2.80 $\pm$ 0.20	2.92 $\pm$ 0.19
P, %	3.03 $\pm$ 0.15 <sup>*a</sup>	1.27 $\pm$ 0.12 <sup>a</sup>	0.55 $\pm$ 0.01	0.48 $\pm$ 0.02
Cl, %	0.93 $\pm$ 0.01 <sup>*</sup>	0.38 $\pm$ 0.01 <sup>a</sup>	1.08 $\pm$ 0.05	0.93 $\pm$ 0.04
K, %	1.72 $\pm$ 0.09 <sup>a</sup>	2.22 $\pm$ 0.20 <sup>a</sup>	2.89 $\pm$ 0.12	3.26 $\pm$ 0.13
Ca, %	0.48 $\pm$ 0.03 <sup>*a</sup>	0.20 $\pm$ 0.03	0.13 $\pm$ 0.01	0.14 $\pm$ 0.03
Zn, mg kg <sup>-1</sup>	37.1 $\pm$ 4.6 <sup>a</sup>	45.4 $\pm$ 4.5 <sup>a</sup>	25.9 $\pm$ 1.9	30.6 $\pm$ 2.4
Br, mg kg <sup>-1</sup>	29.2 $\pm$ 0.4 <sup>*</sup>	6280 $\pm$ 106 <sup>a</sup>	28.6 $\pm$ 1.4 <sup>*</sup>	769 $\pm$ 10
I, mg kg <sup>-1</sup>	1.49 $\pm$ 0.10	1.52 $\pm$ 0.08 <sup>a</sup>	1.31 $\pm$ 0.07 <sup>*</sup>	0.64 $\pm$ 0.05
Nd, mg kg <sup>-1</sup>	0.29 $\pm$ 0.02 <sup>*</sup>	984 $\pm$ 13 <sup>a</sup>	0.40 $\pm$ 0.15 <sup>*</sup>	19.5 $\pm$ 0.7
	Leaves (water-grown plants)		Leaves (soil-grown plants)	
Na, %	0.64 $\pm$ 0.05 <sup>a</sup>	0.60 $\pm$ 0.02 <sup>a</sup>	0.29 $\pm$ 0.01	0.28 $\pm$ 0.02
Mg, %	0.20 $\pm$ 0.04 <sup>a</sup>	0.14 $\pm$ 0.02 <sup>a</sup>	0.40 $\pm$ 0.02	0.38 $\pm$ 0.01
P, %	3.17 $\pm$ 0.25 <sup>a</sup>	2.60 $\pm$ 0.18 <sup>a</sup>	1.1 $\pm$ 0.1	1.2 $\pm$ 0.1
Cl, %	1.05 $\pm$ 0.02 <sup>*a</sup>	0.28 $\pm$ 0.02 <sup>a</sup>	1.53 $\pm$ 0.05	1.28 $\pm$ 0.02
K, %	4.07 $\pm$ 0.21 <sup>a</sup>	4.03 $\pm$ 0.18 <sup>a</sup>	7.43 $\pm$ 0.25	6.72 $\pm$ 0.16
Ca, %	0.26 $\pm$ 0.04 <sup>a</sup>	0.37 $\pm$ 0.07 <sup>a</sup>	0.055 $\pm$ 0.002	0.054 $\pm$ 0.001
Zn, mg kg <sup>-1</sup>	37.4 $\pm$ 2.5 <sup>a</sup>	42.8 $\pm$ 2.5 <sup>a</sup>	48.5 $\pm$ 1.5	56.1 $\pm$ 1.9
Br, mg kg <sup>-1</sup>	86.1 $\pm$ 4.0 <sup>*a</sup>	3351 $\pm$ 50 <sup>a</sup>	31.0 $\pm$ 1.0 <sup>*</sup>	1131 $\pm$ 43
I, mg kg <sup>-1</sup>	0.15 $\pm$ 0.04 <sup>a</sup>	0.22 $\pm$ 0.04	0.25 $\pm$ 0.02	0.23 $\pm$ 0.02
Nd, mg kg <sup>-1</sup>	0.29 $\pm$ 0.01 <sup>*</sup>	63.2 $\pm$ 3.0 <sup>a</sup>	0.29 $\pm$ 0.02 <sup>*</sup>	0.54 $\pm$ 0.04

\* Differences between plants grown in control and in contaminated by bromide media are statistically significant (P<0.05). <sup>a</sup> Differences between plants grown in water and in soil are statistically significant (P<0.05).

**Table 2** Ratios of K/Na and Ca/Mg in wheat seedlings grown in water and soil.

1 – non-contaminated media, 2 – contaminated by bromide of Nd media

Roots				
	Water-grown		Soil-grown	
	1	2	1	2
K/Na	0.48	0.69	10.7	12.1
Ca/Mg	1.2	0.38	0.05	0.05
Leaves				
K/Na	6.4	6.7	26	24
Ca/Mg	1.3	2.7	0.14	0.14